

# Resolution of inflammation pathways in preeclampsia—a narrative review

Luiza Oliveira Perucci<sup>1,2</sup> · Mário Dias Corrêa<sup>3</sup> · Luci Maria Dusse<sup>1,2</sup> · Karina Braga Gomes<sup>1,2</sup> · Lirlândia Pires Sousa<sup>1,2</sup>

Published online: 8 April 2017  
© Springer Science+Business Media New York 2017

**Abstract** Preeclampsia (PE) is one of the leading causes of maternal morbidity and mortality worldwide. This disease is believed to occur in two stages with placental dysfunction in early pregnancy leading to maternal clinical findings after 20 weeks of gestation, as consequence of systemic inflammation, oxidative stress, and endothelial dysfunction. Much evidence suggests that PE women display an overshooting inflammatory response throughout pregnancy due to an unbalanced regulation of innate and adaptive immune responses. Recently, it has been suggested that dysregulation of endogenous protective pathways might be associated with PE etiopathogenesis. Resolution of inflammation is an active process coordinated by mediators from diverse nature that regulate key cellular events to restore tissue homeostasis. Inadequate or insufficient resolution of inflammation is believed to play an important role in the development of chronic inflammatory diseases, like PE. In this narrative review, we discuss possible pro-resolution pathways that might be compromised in PE women, which could be targets to novel therapeutic strategies in this disease.

**Keywords** Preeclampsia · Inflammation · Resolution · Pro-resolving mediators

## Introduction

Preeclampsia (PE) has been defined as a new onset of hypertension and either proteinuria or end-organ dysfunction at gestational age  $\geq 20$  weeks as consequence of systemic inflammation, endothelial dysfunction, and oxidative stress [1, 2]. Because PE is a heterogeneous disease, different classifications based on severity (mild PE/severe PE) and onset of clinical symptoms (early PE  $< 34$  weeks/late PE  $\geq 34$  weeks; pre-term PE  $< 37$  weeks/term PE  $\geq 37$  weeks) have been proposed [3, 4]. It is widely accepted that early PE and late PE have different clinical features, prognosis, and probably distinct etiopathogenesis [5, 6].

Traditionally, a “two-stage” theory of PE etiopathogenesis has been considered. According to this theory, an abnormal spiral artery remodeling in early pregnancy causes placental hypoxia (*stage 1*) and the ischemic placenta releases large amounts of soluble factors, such as reactive oxygen species, pro-inflammatory cytokines, and anti-angiogenic factors, into the maternal circulation, which lead to the clinical manifestations and complications of the disease (*stage 2*) [7, 8]. Another paradigm has been recently proposed by Ahmed and Ramma [9], in which they use a metaphor to compare normotensive pregnancy as a car with accelerators and functioning brakes. The “accelerators” represent inflammation, oxidative stress, and an anti-angiogenic state, while the “brakes” are the endogenous protective pathways. According to this theory, PE manifests when the braking systems fail and the accelerators cannot be stopped in early pregnancy. In their review, Ashmed and Ramma focused on the carbon monoxide, hydrogen sulfide, and nitric oxide pathways. These gases have been

✉ Lirlândia Pires Sousa  
lipsousa72@gmail.com

<sup>1</sup> Departamento de Análises Clínicas e Toxicológicas, Faculdade de Farmácia, Universidade Federal de Minas Gerais, Avenida Antônio Carlos, 6627, Pampulha, Belo Horizonte, Minas Gerais 31270-901, Brazil

<sup>2</sup> Programa de Pós-Graduação em Análises Clínicas e Toxicológicas, Universidade Federal de Minas Gerais, Avenida Antônio Carlos, 6627, Pampulha, Belo Horizonte, Minas Gerais 31270-901, Brazil

<sup>3</sup> Departamento de Ginecologia e Obstetrícia, Faculdade de Medicina, Universidade Federal de Minas Gerais, Avenida Professor Alfredo Balena, 190, Santa Efigênia, Belo Horizonte, Minas Gerais 30130-100, Brazil

associated with protective roles, such as regulation of uteroplacental perfusion and inhibition of oxidative stress and inflammation [10–12]. The new paradigm of dysregulated endogenous protective pathways can be combined with the traditional two-stage theory of PE pathogenesis (Fig. 1). Here, we raise the hypothesis of another protective pathway that may be compromised in PE women, the resolution of inflammation pathway.

**Resolution of inflammation**

Acute inflammation is usually a self-limited response that can be triggered by infectious or sterile injury and has the physiological purpose to restore tissue homeostasis [13]. Successful resolution of inflammation is an active and highly regulated process that evolves several cellular and biochemical events [14, 15]. During this process, the production of anti-inflammatory/pro-resolving factors prevails over the production of pro-inflammatory mediators. However, inflammation and resolution are not isolated events. In fact, they continuously overlap because pro-inflammatory signals can induce anti-inflammatory and pro-resolving signals aiming to temper inflammation [16, 17].

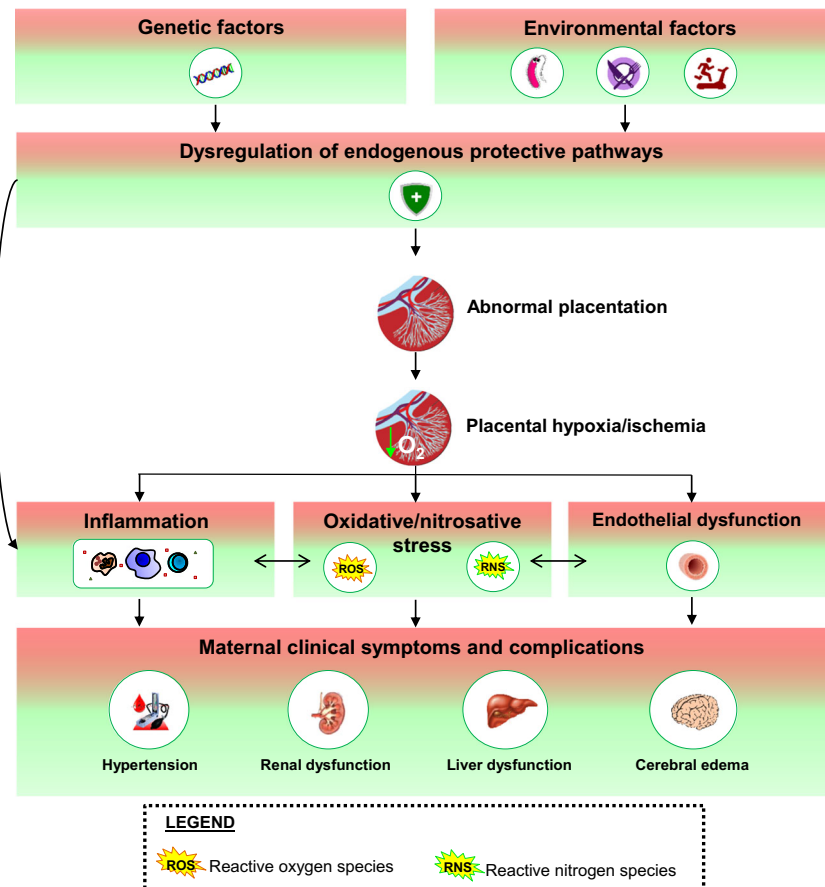
In recent years, endogenous pro-resolution mediators from diverse nature have been identified, including proteins/

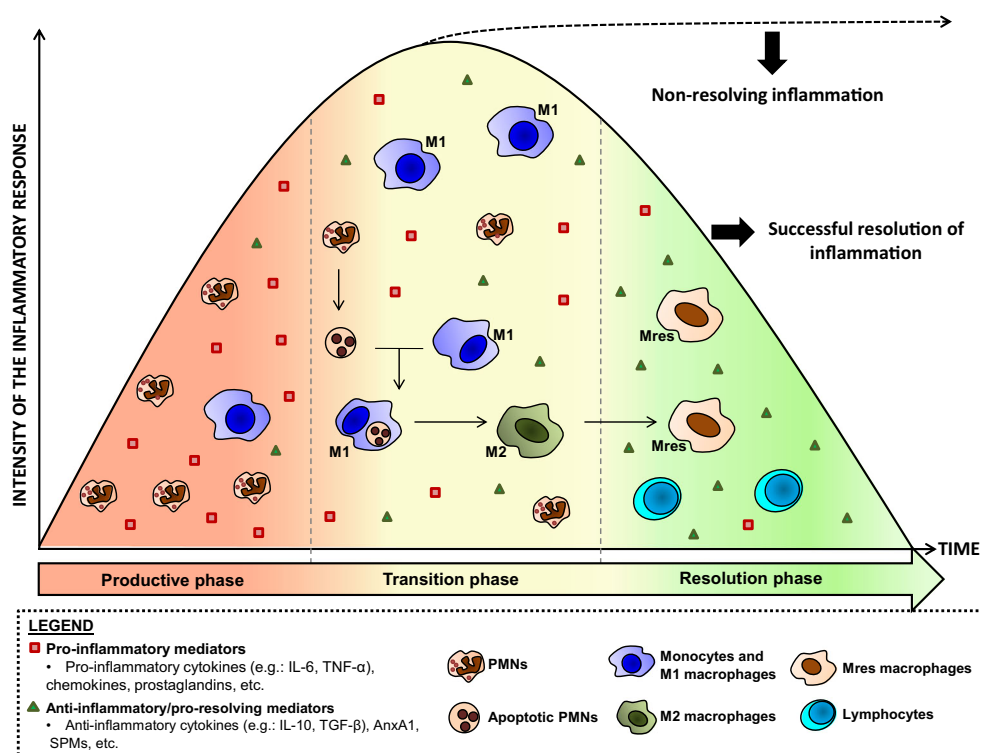
peptides, specialized pro-resolving lipid mediators, gaseous mediators, protease inhibitors, and neuromodulators [16, 18, 19]. They inhibit further leukocyte recruitment, induce neutrophil apoptosis, and enhance the efferocytosis of apoptotic neutrophils by macrophages, thus acting as brakes for the inflammatory response. They are also able to switch macrophages from pro-inflammatory (M1) to anti-inflammatory and pro-resolving phenotypes (M2 and Mres), drain non-apoptotic leukocytes to lymph nodes, and participate in tissue repair/healing mechanisms [14–16, 20]. Figure 2 shows the key steps of successful resolution of an inflammatory process.

Inflammation may become chronic and lead to further tissue damage if resolution process fails. Dysfunctional resolution of inflammation can occur due to decreased synthesis of pro-resolving mediators and receptors, altered receptors conformation, and increased inactivation of pro-resolving mediators. Inadequate amount or action of pro-resolving mediators can lead, for example, to persistent recruitment and survival of neutrophils, failure to reprogram macrophage phenotype, and ineffective clearance of apoptotic neutrophils. In this sense, if the inflammatory stimulus is too high, it would be necessary a higher production of anti-inflammatory/pro-resolving molecules to neutralize the overshooting inflammation.

In a recent review, Fullerton and Gilroy proposed that resolution could be a bridge between innate and adaptive

**Fig. 1** The combination of “the two-stage” and “the accelerator and brake” theories might explain PE etiopathogenesis. This schematic diagram illustrates the sequential events involved in PE etiopathogenesis. Genetic and environmental factors disrupt endogenous protective pathways, leading to inadequate invasion of uterine spiral arteries by placental trophoblasts and a failure of physiological transformation of uterine spiral arteries. This results in placental hypoxia/ischemia. The dysfunctional placenta releases large amounts of soluble factors into the maternal circulation, which lead to generalized inflammation, oxidative stress/nitrosative stress, and endothelial dysfunction, events that are interconnected and precede PE clinical symptoms and complications. Alternatively, dysregulation of endogenous protective pathways can directly cause inflammation, oxidative stress, and endothelial dysfunction





**Fig. 2** Success and failure in resolution of inflammation. The productive phase of the inflammatory response is characterized by a significant influx of PMN cells to the inflamed tissue and by an increased generation of pro-inflammatory mediators by endothelial cells, migrated and resident immune cells. As the inflammatory response progresses (transition phase), there is a switch in the production of pro-inflammatory mediators to anti-inflammatory mediators and a reduction of PMN cell migration that parallel an increase in the influx of mononuclear cells. In addition, pro-inflammatory signals induce PMN cell apoptosis and macrophage phagocytosis of apoptotic PMN cells (efferocytosis). During this process, pro-inflammatory macrophages (M1) alter their phenotype to anti-inflammatory macrophages (M2). M2

macrophages have greater ability of efferocytosis and to produce anti-inflammatory/pro-resolving mediators. During the resolution phase of inflammation, the influx of monocytes prevails over the influx of PMN; the synthesis of anti-inflammatory/pro-resolving mediators is increased, while the levels of pro-inflammatory mediators are decreased. Moreover, M2 macrophages are converted into a pro-resolving phenotype (Mres), with greater ability to produce anti-inflammatory/pro-resolving mediators, and lymphocytes repopulate the affected tissue. Collectively, these events lead to successful resolution of acute inflammation. On the other hand, failures in resolution of inflammation pathways lead to persistent inflammation and maladaptive immune responses

immunities. Therefore, unresolved inflammation could lead to maladaptive immune responses, which are commonly associated with chronic inflammatory diseases [21].

## Hypothesis

Embryo implantation, trophoblast invasion of uterine spiral arteries, and labor are inflammatory events. Therefore, inflammation is necessary to successful reproduction [22]. Normotensive pregnancy is characterized by a state of mild/low-grade inflammation, as demonstrated, for example, by increased levels of pro-inflammatory cytokines when compared to the non-pregnant state [23]. Innate immune responses are upregulated in normotensive pregnant women, while adaptive immune responses are modulated in order to maintain maternal immune tolerance to the

fetal allograft. By contrast, innate immune responses are even more activated and adaptive immune responses are dysregulated in PE [24]. Indeed, there is a shift from T helper (Th)2/regulatory T cell responses in normotensive pregnant women to a predominant Th1/Th17 immunity in PE women [25]. Furthermore, there are evidences of placental M2 macrophage polarization in normotensive pregnancy and a predominant M1 phenotype in PE [26, 27]. Consequently, PE women display an overshooting inflammatory response throughout pregnancy [23].

Most studies of the literature have focused on the accelerators of the inflammatory response in PE. Here, we propose that the exaggerated inflammatory response seen in this disease may result from failures in a “braking system” called resolution of inflammation. If so, unresolved inflammation may account for maladaptive immune responses in PE women.

## Methods

First, we performed a screening on PubMed database results through reading of titles and abstracts about pro-resolving mediators previously described in general works [16, 18, 19]. We used as key terms the specific pro-resolving mediator and preeclampsia. The pro-resolving mediators that had been studied in the context of PE according to this research were annexin A1, galectins, chemerin, lipoxin A4, nitric oxide, hydrogen sulfide, carbon monoxide, acetylcholine, netrin-1, and protease inhibitors. The final selection was based on full reading of each preselected article. Original research articles were included if they addressed these pro-resolving mediators in the context of PE (human disease and animal models). Original articles about pro-resolving mediators in other inflammatory diseases and general review articles were also included to provide a background on the role of these mediators. Articles that had not focused on these issues were excluded.

## Results

This review included a total of 225 articles published between 1985 and 2017. The main conclusions obtained from them are described in the next subsections.

### Annexin A1

Annexin A1 (AnxA1) is a 37-kDa glucocorticoid-regulated protein that elicits anti-inflammatory/pro-resolving effects through binding to formyl peptide receptor type 2/lipoxin A4 receptor (FPR2/ALXR). These effects lie within AnxA1 N-terminal domain and include inhibition of neutrophil migration to inflamed tissues, induction of neutrophil apoptosis, stimulation of macrophage efferocytosis of apoptotic neutrophils, and induction of macrophage reprogramming to a pro-resolving phenotype [28, 29]. AnxA1 was first recognized by its capacity to inhibit phospholipase A2 activity and the generation of eicosanoids, but subsequent studies revealed that this protein exerts a wider range of actions. It has been suggested that AnxA1 mediates part of neuroendocrine responses of the glucocorticoids, particularly in the hypothalamic-pituitary-adrenocortical axis. In addition, experimental data indicate that AnxA1 may be implicated in processes regulating pregnancy, lactation, and fetal development [30, 31].

Altered AnxA1 synthesis might be involved in the pathogenesis of chronic inflammatory diseases, like asthma [32]. Of importance, intact AnxA1 (37 kDa) can be cleaved in its N-terminal domain by proteases, such as neutrophil elastase, generating various fragments that are believed to be inactive or pro-inflammatory [33]. Indeed, increased levels of AnxA1 cleavage products (e.g., 33 kDa) have been reported in inflammatory samples [34]. Moreover, FPR2/ALXR decreased

expression or altered receptor conformation can impair AnxA1 to regulate inflammation [35, 36].

Previously, Perucci et al. investigated AnxA1 in PE and found increased plasma levels when compared to normotensive pregnancy [37]. The increased concentration of AnxA1 combined with an overwhelming inflammatory response suggests a failure in this resolution pathway in PE, which could be a consequence of decreased expression of FPR2/ALXR [38, 39] or presence of anti-AnxA1 auto-antibodies [40]. Considering that neutrophilia is a common feature in PE women and that neutrophil elastase is increased in their plasma and placenta [41–43], it is also plausible to hypothesize that AnxA1 cleavage could interfere with its actions. Although AnxA1 expression has been studied in placental tissues [44], the differential expression of its intact and cleaved forms in preeclamptic and normotensive pregnancies has not been determined, a matter under investigation in our group.

### Galectins

Galectins are  $\beta$ -galactoside-binding proteins that were initially known to mediate developmental processes, including tissue organization and embryo implantation [45]. Further research indicated that galectins are secreted in response to inflammatory signals and cellular damage, acting as pattern recognition receptors, immunomodulators, or damage-associated molecular patterns in innate and adaptive immune responses [46, 47]. Galectins are thought to modulate intracellular signaling pathways in immune cells due to their ability to induce the aggregation of specific cell-surface glycoreceptors [48, 49]. Thereby, galectins may elicit pro-resolving effects, as described below. Emerging evidences also suggest that galectins are capable of triggering platelet activation and inducing angiogenesis [50, 51]. Here, we give a general overview on the role of galectin-1 (Gal-1) and galectin-13 (Gal-13), the most studied galectins in PE.

#### *Galectin-1*

It has been suggested that Gal-1 plays a role in maternal-fetal tolerance, which is thought to be impaired in PE women. Blois et al. reported that Gal-1 deficient (*LGALS1*<sup>-/-</sup>) mice had increased fetal loss when compared to wild-type mice, an effect that was prevented by the treatment with recombinant Gal-1. According to their results, Gal-1 restored maternal immune tolerance by promoting the expansion of IL-10-secreting regulatory T cells [52]. This data was corroborated by the study of van der Leij et al. [53]. Gal-1 might also improve maternal-fetal tolerance by inducing the apoptosis of activated CD8<sup>+</sup> T cells, Th1 cells, and Th17 CD4<sup>+</sup> cells [54]. Other



immunomodulatory actions of Gal-1 have been proposed. For instance, Rostoker et al. showed that Gal-1 induced 12/15-lipoxygenase expression (lipoxin A4 synthesizing enzyme; see the “**Lipoxin A4**” section) in murine macrophages and promoted their conversion into a pro-resolving phenotype [55].

Some works have demonstrated that the gene expression of Gal-1 was upregulated in placentas from PE women compared with normotensive pregnant women [56–58]. Interestingly, *LGALS1*-knockout dams develop PE symptoms. However, when stratifying PE women according to the onset of clinical symptoms, early PE women showed lower placental expression of Gal-1 than pregnant controls, while an opposite finding was reported for late PE women [58]. It has been proposed that the decreased expression of Gal-1 in early PE could be associated with placental dysfunction, whereas its overexpression might be a compensatory mechanism to attenuate inflammation in late PE [59]. In addition, the circulating levels of Gal-1 may reflect its placental expression in late PE, but not in early PE. Accordingly, Freitag et al. reported increased serum levels of Gal-1 in late PE when compared with early PE and normotensive pregnancies, but no difference was found between early PE women and normotensive women [58]. However, when both clinical forms were included in the same cohort, the serum levels of Gal-1 seemed to be similar between patients and controls [60]. Pregnant women in the second trimester of pregnancy who developed PE also showed lower levels of Gal-1 than healthy pregnant women, indicating that Gal-1 might be an early predictor of PE [58].

Gal-1 seems to be differentially expressed in cells/tissues from women with PE. Gal-1 is downregulated in T and natural killer cells in PE when compared with these cells from normotensive pregnancy, while no difference was detected in Gal-1 messenger RNA (mRNA) expression in decidual samples between these pregnant groups [58, 60]. The decreased expression of Gal-1 expression in these immune cells can be associated with maternal-fetal intolerance and exacerbated inflammatory response in PE women, as discussed above.

### *Galectin-13*

Gal-13 is a galectin uniquely expressed in the placenta, mainly in the syncytiotrophoblast, and it is released from the placenta into the maternal circulation [61]. In vitro studies suggest that Gal-13 participates in the morphological differentiation of the cytotrophoblast into the syncytiotrophoblast [62, 63]. In addition, it has been demonstrated that Gal-13 is able to induce the apoptosis of activated human CD3<sup>+</sup> T cells [64]. Interestingly, phagocytosed Gal-13 immunopositive deposits in immune cells coincided with zones of apoptotic and necrotic immune cells in Kliman et al. study [65]. These data indicate that Gal-13 might participate in placentation and in maternal adaptive immune responses at the maternal-fetal interface. Considering

that these processes are impaired in PE women, it can be hypothesized that Gal-13 is involved in the pathogenesis of the disease.

Gal-13 placental-specific expression makes it a promising biomarker for PE early prediction. Indeed, Gal-13 protein and mRNA content in blood and placenta are decreased in the first trimester of gestation in women who developed PE, especially in the early clinical form [66–71], and this could be associated with single-nucleotide polymorphisms in the *LGALS13* gene [72]. Moreover, combining Gal-13 with background risk factors, other serum biomarkers and physical parameters increase the accuracy of predicting PE [73]. Low serum levels of Gal-13 in early gestation may lead to impaired placentation and maternal immune intolerance to the fetus [65, 74].

It has been demonstrated that the serum levels of Gal-13 increase throughout normotensive pregnancy and that preterm PE women have higher serum levels of Gal-13 than preterm controls [66]. It was proposed that the increase in maternal serum concentration of Gal-13 during the third trimester of gestation in PE women is a consequence of augmented placental shedding of microvesicles containing Gal-13, and this could be a compensatory mechanism aiming to restore homeostasis [66]. Nevertheless, both decreased and increased expressions of placental Gal-13 have been reported in PE women [66, 75].

### **Chemerin**

Chemerin is an adipocyte-secreted protein originally identified as the natural ligand of chemR23 receptor, which is implicated in several biological processes, such as adipogenesis, glucose homeostasis, and immune cell migration [76]. It has been suggested that chemerin is abundantly expressed in stromal cells and in extravillous trophoblast cells, but not in decidual endothelial cells in early pregnancy [77]. Moreover, chemerin may stimulate angiogenesis and the accumulation of natural killer cells at maternal-fetal interface, and these immune cells have been implicated in uterine spiral artery remodeling [77–79]. Thus, chemerin can be involved in placental development, which is impaired in PE women. Chemerin also acts as a chemoattractant for dendritic cells [80]. Several lines of evidence indicate crucial roles for both natural killer and dendritic cells in the modulation of adaptive immune responses [81, 82]. Based on these data, it can be admitted that chemerin may contribute to maternal-fetal tolerance, but more studies are necessary to clarify this issue.

Fragments with distinct inflammatory actions can be generated after chemerin C-terminal proteolytic processing, depending on the types of proteases predominating in the microenvironment [80, 83]. Some chemerin fragments can induce the chemotaxis of immune cells, in particular dendritic cells, macrophages, and natural killer cells, toward inflammatory sites, thus contributing to the onset of inflammation. By

contrast, other fragments can inhibit the synthesis of pro-inflammatory mediators. In addition, the activation of the chemerin/chemR23 axis may increase the non-phlogistic phagocytosis of apoptotic cells by macrophages, and inhibit neutrophil activation and influx to inflammatory sites, thus promoting the resolution of inflammation. Therefore, chemerin-derived peptides may play a role both in initiation and in resolution of the inflammatory response [80].

It has been shown that the serum levels of chemerin increase throughout normotensive pregnancy [84, 85]. Some studies have reported increased circulating levels of chemerin, as well as increased mRNA and protein expressions in placentas from PE women when compared to normotensive pregnant women [86–88]. Higher levels of chemerin were detected in the first trimester of gestation in women who developed PE, were associated with disease severity, and remained significantly higher 6 months after delivery in former PE women compared with controls [87–89]. Moreover, there is a positive correlation among chemerin levels, pro-inflammatory mediators, and blood pressure [86–89]. However, these studies did not specify the types of chemerin-derived peptides quantified. Hence, their role in PE pathogenesis remains unclear.

### Specialized pro-resolving lipid mediators

The polyunsaturated fatty acids omega-6 and -3 are substrates for the biosynthesis of lipoxins (LXs), maresins, resolvins, and protectins, which are collectively called *specialized pro-resolving lipid mediators* (SPMs). Prostaglandins and leukotrienes are lipid mediators that play pivotal roles in the initiation of the inflammatory response, while SPMs attenuate inflammation and contribute to its timely resolution [90]. Curiously, aspirin induces the endogenous synthesis of LX 15-epimers [91]. Endogenous LXs and their epimers have been shown to counter-regulate inflammation in a variety of experimental models of inflammatory diseases. They down-regulate pro-inflammatory mediators' synthesis (including prostaglandins and leukotrienes), inhibit neutrophil infiltration, induce macrophage efferocytosis of apoptotic neutrophils, and stimulate interleukin (IL)-10 production [90, 92]. Furthermore, LXs may modulate other biological actions, such as angiogenesis, airway smooth muscle function, and activity of neuronal ion channels that convey nociceptive signals [93–95]. In this sense, LXs and other SPMs may contribute to resolution of both inflammation and pain [94].

#### *Lipoxin A4*

Lipoxin A4 (LXA4) is an eicosanoid synthesized from arachidonic acid, an omega-6 derivate, through the metabolism of lipoxygenase enzymes [96]. LXA4 interacts with FPR2/ALXR receptor, which also binds to AnxA1 [97]. An *in vitro* study showed that LXA4 inhibited the production of

IL-1 $\beta$  by monocytes from severe PE women in a dose-dependent manner [98]. In another experiment, 15-epi-LXA4 reduced neutrophil-endothelium cell adhesion triggered by PE plasma [99]. Lin et al. administrated an LXA4 analogue in low-dose-endotoxin-treated pregnant rats and found that it attenuated inflammation and PE symptoms [100]. These experimental data suggest protective roles for LXA4 and its analogues in the disease.

Three works showed higher circulating levels of LXA4 in PE women compared to normotensive pregnant women [39, 101, 102]. However, an opposite finding has also been reported [38]. Different studied populations or methodologies to quantify LXA4 might have contributed to these divergent results. Interestingly, LXA4 plasma levels correlated with maternal blood pressure, white blood cell count, and C-reactive protein levels in Perucci et al. study [102]. Similar to AnxA1 discussion, LXA4 inefficiency to resolve inflammation could be a consequence, for example, of the decreased expression of FPR2/ALXR and/or increased LXA4 inactivation. However, these hypotheses remain to be investigated.

### Gaseous mediators

Nitric oxide, hydrogen sulfide, and carbon monoxide are the most studied gaseous mediators, and, for many years, only their toxicity was known [103]. Recently, they have been implicated in key physiological functions, such as angiogenesis, inflammation, and vascular tone regulation [104, 105]. They also participate in trophoblast invasion and in spiral artery remodeling [106]. Experimental studies have demonstrated that these gases act as anti-inflammatory mediators at low concentrations, promoting resolution of inflammation, but exert pro-inflammatory and damaging effects at high concentrations [12]. In line with this data, altered production or signaling of gaseous mediators has been reported in inflammatory diseases, like atherosclerosis and arthritis [107, 108].

#### *Nitric oxide*

Nitric oxide (NO) is synthesized by the conversion of L-arginine to L-citrulline by one of the following three isoforms of nitric oxide synthase (NOS): neuronal, endothelial (eNOS), or inducible (iNOS). NO acts as a vasodilatory molecule by inducing cyclic guanosine monophosphate (cGMP) synthesis [109]. However, NO can act by cGMP-independent pathways to regulate other mechanisms, such as leukocyte apoptosis [110, 111]. NO may have pro- or anti-inflammatory actions depending on the concentration used in the experiment, the delivery method, and the system/disease model studied [112]. Low amounts of NO inhibit the synthesis of pro-inflammatory cytokines and reduce leukocyte-endothelium adhesion and transmigration to inflamed tissues, while high levels of NO increase vascular permeability and leukocyte migration [113,

[114]. NO might also play a role in resolution of inflammation since it induces neutrophil, but not macrophage, apoptosis [115].

In normal pregnancy, NO and cGMP biosynthesis are increased due to eNOS upregulation. In addition, the biosynthesis of asymmetrical dimethylarginine (ADMA), a competitive inhibitor of NOS, is reduced. These events are important to regulate peripheral and placental bed vascular resistances, angiogenesis, platelet adhesion/aggregation, and trophoblast invasion [116]. On the other hand, most studies have reported decreased activity of placental iNOS and eNOS and increased levels of ADMA in PE, but data on NO levels are inconsistent [117–119]. Moreover, increased levels of ADMA in the first trimester of pregnancy may predict PE [120]. Additionally, ADMA increase seems to be more prominent in early severe PE than in late severe PE and eNOS polymorphisms may influence the onset time of the disease [121, 122].

Other mechanisms may interfere with NO signaling in PE. It has been demonstrated that polymorphisms in the transcription factor STOX1 gene are associated with maternal susceptibility to PE and that the overexpression of placental STOX1 induces a PE-like syndrome in mice [123, 124]. According to the study of Doridot et al., placentas overexpressing STOX1 showed high concentration of reactive nitrogen species (RNS). They proposed that RNS could be rapidly generated in the placenta through the association of NO with reactive oxygen species (ROS) [125]. Therefore, the overexpression of STOX1 could decrease NO bioavailability in endothelial cells, preventing the protective actions of this gaseous mediator in vascular tone and inflammation and also contributing to oxidative and nitrosative stress in PE women [11]. Further evidence suggested a risk allele (Y153H) in STOX1 gene that might be associated with a less invasive trophoblast phenotype [126]. Accordingly, a previous study showed that the ability of trophoblasts to remodel uteroplacental arteries depended on NO produced by extravillous trophoblasts in guinea pig pregnancy [127].

Considering that NO interferes with several pathways that are known to be compromised in PE women, such as vascular tone, inflammation, and oxidative/anti-oxidative status, impaired bioavailability and/or action of this gaseous mediator may be associated with the pathogenesis of the disease. In this sense, the therapeutical potential of drugs that enhance NO availability, inhibit cGMP degradation, or reduce ADMA levels has been investigated in vitro and in experimental models of PE. PE was mimicked by inducing reduced uterine perfusion pressure (RUPP) model and the overexpression of the anti-angiogenic molecule soluble fms-like tyrosine kinase (sFlt1) and by administrating the NOS inhibitor  $N^G$ -nitro-L-arginine methyl ester (L-NAME) in rodents [128]. Although some of these findings seem promising, there is insufficient clinical evidence to use these drugs for PE treatment or prevention [128, 129].

### Hydrogen sulfide

Endogenous hydrogen sulfide ( $H_2S$ ) is primarily synthesized by the conversion of L-cysteine or homocysteine by two enzymes, which are cystathionine  $\beta$ -synthase (CBS) and cystathionine  $\gamma$ -lyase (CSE) [130].  $H_2S$  is a vasodilatory mediator, an effect that can be mediated through eNOS activation and NO production [131].  $H_2S$  exert anti-oxidant effects on cells and it seems to be cytoprotective at low concentrations, while higher  $H_2S$  exposure favors oxidative stress and cell apoptosis [132]. As for NO,  $H_2S$  role in inflammation is complex and not fully elucidated.  $H_2S$  might act as a pro-inflammatory mediator, as demonstrated in experimental sepsis [133, 134], or as an anti-inflammatory/pro-resolving molecule, for example, in gastrointestinal inflammation [135]. Evidences suggest that  $H_2S$  promotes resolution of inflammation by inducing neutrophil apoptosis, M2 macrophage polarization, and clearance of apoptotic neutrophils by macrophages [136–138]. Recently, it has been suggested that part of  $H_2S$  anti-inflammatory/pro-resolving effects are mediated by AnxA1 [139].

$H_2S$  has been implicated in placental vascular development and function due to its pro-angiogenic and vasodilatory activities [131, 140]. Both  $H_2S$ -synthesizing enzymes CBS and CSE are expressed in human placenta during normal pregnancies [141], but the studies on their expression in PE have conflicting results. Wang et al. reported a downregulation of CSE (mRNA and protein) in placentas from PE women [142]. Moreover, placentas with abnormal Doppler have increased expression of microRNA-21, which negatively regulates CSE expression [143]. By contrast, in Holwerda et al. study, mRNA levels of CSE in the placenta were unchanged, while mRNA levels of CBS were decreased in early PE [144]. These divergent results might be attributed to differences in the studied clinical forms of PE [128]. Furthermore, Wang et al. reported decreased plasma levels of  $H_2S$  in PE women [142]. Abnormal synthesis of  $H_2S$  could contribute to endothelial dysfunction, oxidative stress, and overwhelming inflammation observed in PE women [128].

$H_2S$ -based therapies have been studied in animal models of PE and in human disease. The administration of a slow-releasing  $H_2S$ -generating compound (GYY4137) ameliorated PE-like symptoms induced by the treatment with an inhibitor of  $H_2S$  synthesis (DL-propargylglycine) [142]. However, oral administration of an  $H_2S$  donor (N-acetylcysteine) to severe early PE women did not improve maternal outcomes [145]. More studies should be conducted in order to evaluate the therapeutical potential of  $H_2S$ -releasing compounds in PE.

### Carbon monoxide

Heme oxygenase (HO) enzymes convert heme to biliverdin, free iron, and carbon monoxide (CO) in the endoplasmatic

reticulum. HO enzymes exist as inducible (HO-1) and constitutive (HO-2) isoforms [146]. HO-CO system regulates many biological processes, such as vascular tone, oxidant-anti-oxidant status, and platelet aggregation. Further, CO acts as a signaling molecule in the neuronal system, where it regulates the release of neurotransmitters. Like NO and H<sub>2</sub>S, CO is toxic at high concentrations but has cytoprotective actions at low concentrations [147, 148]. It has been suggested that part of the protective and deleterious effects of CO are due to its ability to regulate different types of ion channels [149]. Most studies have reported counter-regulatory actions for CO in inflammatory responses. Low CO exposure inhibits neutrophil-endothelial adhesion and transmigration to inflamed tissues, suppresses the production of pro-inflammatory cytokines, promotes neutrophil apoptosis, and enhances macrophage efferocytosis of apoptotic neutrophils [150–153]. Moreover, CO accelerates resolution of inflammation by shifting the lipid profile in the inflammatory milieu [153].

Both HO-1 and HO-2 are expressed in human placenta [154]. During pregnancy, CO regulates perfusion and oxidant-anti-oxidant status within placental tissues, as well as spiral artery transformation [154–156]. Adequate expression of HO might also be important to maintain maternal-fetal tolerance [157]. Considering the importance of CO in regulating multiple processes during pregnancy, alterations in the HO-CO system in PE would be expected. Indeed, PE women seem to have lower CO breath levels and carboxyhemoglobin concentration in the umbilical cord blood than normotensive pregnant women [158, 159]. These data are in line with the observation that CO exposure in cigarette smoke decreases the risk of developing PE [160, 161]. However, HO placental expression during PE is not clear. Either decreased, increased, or unchanged, placental expressions of HO-1 and HO-2 have been reported in PE women [162–166].

Compounds that induce HO expression have been studied in experimental and in human PE [128]. A recent work showed that pravastatin treatment stabilized blood pressure, proteinuria, and serum uric acid levels in severe PE women [167]. These effects seem to be partially mediated by upregulating HO-1 placental expression [168]. McCarthy et al. studied rosiglitazone effects using the RUPP rat model of PE and found that this drug prevented the development of disease-like symptoms via HO-dependent pathway [169]. Moreover, CO application at low doses prevented hypertension and proteinuria in adenovirus sFlt1 PE-like mouse model [170]. In conclusion, the administration of the CO or HO-inducing agents might be beneficial for treating or preventing PE, but further investigation is necessary.

## Neuromodulators

### *Acetylcholine*

Cholinergic neurons release acetylcholine (ACh), a neurotransmitter known to regulate skeletal, smooth, and cardiac muscle contractions. ACh also acts as neuromodulator in the central nervous system, where it alters neural excitability, synaptic transmission, and plasticity, thus interfering with learning, memory, and mood [171, 172]. ACh can induce endothelium vasodilatation through NO-dependent and independent mechanisms, for example, by inducing the production of prostaglandins [173]. Indeed, it has been demonstrated that pharmacological administration of ACh reduces blood pressure in rats [174]. These data suggest that ACh-deficient synthesis or action may be associated with PE pathogenesis.

Studies on the role of neural reflexes in inflammation and immunity are recent. It has been demonstrated that ACh binding to  $\alpha$ 7-nicotinic receptors in macrophages inhibits the synthesis and the release of pro-inflammatory cytokines [175–178]. Alternative anti-inflammatory cholinergic mechanisms have been proposed. For instance, nicotine (a cholinergic agonist drug) attenuates inflammation by upregulating the expression of HO-1 in macrophages [179, 180]. Other anti-inflammatory and pro-resolving effects of nicotine include inhibition of neutrophil migration and stimulation of its apoptosis [181, 182]. Moreover, ACh receptor activation by nicotine enhances macrophage phagocytosis and protects M2 macrophages from apoptosis [183, 184]. The role of this neural pathway in controlling inflammatory responses was further confirmed by studies showing that vagus nerve lesions enhance pro-inflammatory cytokines' production and are associated with non-resolving inflammation [176, 185, 186]. Accordingly, chronic inflammatory conditions, such as inflammatory bowel disease, have decreased vagus nerve function [176].

Yang et al. reported reduced vagus nerve function in PE women [187]. Thus, ACh-reduced synthesis in these women might contribute to excessive inflammation and hypertension. Accordingly, nicotine binding to ACh receptor suppresses ex vivo placental cytokines' production [188]. In a recent study, nicotine was able to reduce systolic blood pressure in LPS-induced PE rat model [189]. This effect might be associated with nicotine protective effects on the endothelium, as previously demonstrated by Mimura et al. [190]. These data corroborate with the theory that nicotine, and also CO, in cigarette smoke might protect from PE [160, 161]. Some studies have also shown that nicotinic ACh receptors are upregulated in PE women [191, 192], which could be a compensatory mechanism to decreased ACh levels.



## Netrin-1

Netrin-1 was originally described as a laminin-related protein that guides axonal trajectories during the development of central nervous system, by repulsing/abolishing the attraction of neuronal cells expressing the UNC5b receptor [193]. Subsequently, it was implicated in the regulation of various biological processes, including angiogenesis and, recently, inflammation. Netrin-1 suppresses neutrophil trafficking, probably as consequence of the strong expression of UNC5b receptor in these cells [194, 195]. It also inhibits prostaglandin E2 synthesis, regulates Th1/Th2/Th17 cytokines' production, induces M2 polarization, increases apoptotic polymorphonuclear (PMN) cell efferocytosis, and stimulates the endogenous biosynthesis of SPMs [196–199]. In accordance, *in vivo* studies have reported protective functions of netrin-1 in inflammatory conditions [199–202]. Interestingly, Mirakaj et al. found that netrin-1 stimulated the resolution of peritoneal inflammation induced by zymosan via resolvin D1, a pro-resolving lipid mediator [186]. Yang et al. investigated the placental expression of netrin-1 and found that it was downregulated in severe PE women [203]. More studies are needed to understand the association between netrin-1 and inflammation in PE.

## Protease inhibitors

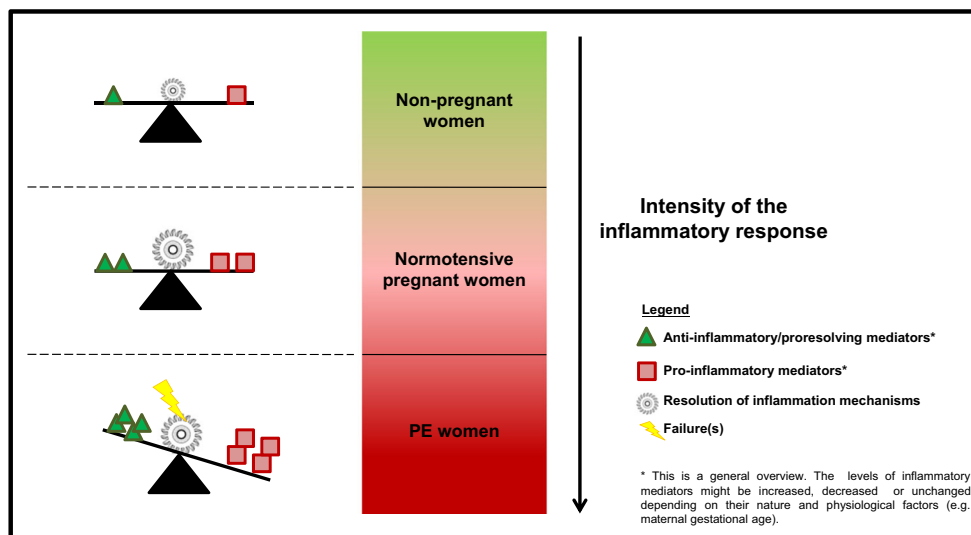
Proteases are enzymes that hydrolyze peptide bonds of proteins, releasing polypeptides or free amino acids. They regulate the activity and the localization of several proteins, modulate the interactions among them, and participate in cellular

signaling events. Currently, proteases are classified based on their mechanisms of catalysis into the following four classes: serine proteases, metalloproteases, aspartic proteases, and cysteine proteases. Their activities are tempered by protease inhibitors or anti-proteases [204, 205]. Proteases are usually upregulated in inflammatory conditions, and defective anti-proteolytic control mechanisms may participate in the pathogenesis of chronic inflammatory diseases, like cystic fibrosis [206, 207]. Thus, protease inhibitors have the potential to be developed as new therapeutic agents for these diseases.

## Metalloproteinase inhibitors

Metalloproteinases are proteolytic enzymes that hydrolyze extracellular matrix components, playing important roles on tissue repair. They participate in extracellular matrix remodeling during trophoblast invasion and in uterine spiral artery transformation. This family of enzymes comprises, among other members, matrix metalloproteinases (MMPs) and membrane-anchored disintegrin metalloproteinases (ADAMs) [208, 209]. Activated metalloproteinases can be regulated by general or specific protease inhibitors (tissue inhibitors of metalloproteinases (TIMPs)) [210].

Several non-matrix substrates for metalloproteinases have been identified, including cytokines, chemokines, and their receptors. Metalloproteinases cleave these substrates in short fragments, altering their bioactions, and, in the case of receptors, interfering with their responsiveness and downstream signaling. Metalloproteinases modulate additional aspects of inflammation, such as integrity of physical barriers,



**Fig. 3** Schematic representation of inflammatory and counter-regulatory mechanisms in non-pregnant women, normotensive pregnant women, and PE women. Healthy non-pregnant women have basal levels of anti-inflammatory/pro-resolving mediators and pro-inflammatory mediators, which are in a state of equilibrium due to functioning resolution of inflammation mechanisms. Normotensive pregnant women show higher levels of pro-inflammatory mediators than non-pregnant women, but the

inflammatory response is mild and controlled, because resolution of inflammation mechanisms are able to adjust properly to this physiological state (increased gear symbol). By contrast, failures in pro-resolving mechanisms probably lead to an exacerbated inflammatory response in PE women, despite the upregulation of some anti-inflammatory/pro-resolving mediators

leukocytes' transmigration, and survival [211, 212]. Metalloproteinases may have pro-inflammatory or anti-inflammatory/pro-resolving actions. For instance, ADAM17, also known as tumor necrosis factor alpha (TNF- $\alpha$ ) converting enzyme, releases the membrane-bound TNF- $\alpha$ , increasing the bioavailability of this pro-inflammatory cytokine. By contrast, ADAM17 sheds TNF- $\alpha$  soluble receptors (sTNFRs) in the circulation, which sequester TNF- $\alpha$ , neutralizing its systemic effects [213]. ADAM17 also prevents neutrophil transmigration through the endothelium by shedding L-selectin from them, without altering monocyte recruitment. In addition, ADAM17 induces neutrophil apoptosis [214, 215].

There is increasing evidence of metalloproteinase/TIMP imbalance in inflammatory diseases, like inflammatory bowel diseases [216]. Metalloproteinases and their inhibitors may also participate in PE pathogenesis. Ma et al. reported that ADAM17 was upregulated in placentas from PE women and induced TNF- $\alpha$  production by placental trophoblasts [217]. Later, they showed that the placental levels of TIMP3 (ADAM17 inhibitor) were decreased in PE women and that TIMP3 downregulation increased TNF- $\alpha$  production by placental trophoblasts [218]. Further reports on increased circulating levels of TNF- $\alpha$  and sTNFRs in PE women corroborated with these findings [219–221], since increased ADAM 17 levels and decreased TIMP3 levels may induce TNF- $\alpha$  release and the consequent shedding of neutralizing sTNFR receptors in the circulation of PE women. Decreased, increased, or similar levels of other metalloproteinases and TIMPs have been described in PE [222]. These discrepancies are probably due to differences in the types of specimens analyzed, gestational age of specimen collection, and quantification methodologies. Therefore, the role of metalloproteinases and their inhibitors in PE pathogenesis requires further investigation.

## Concluding remarks

Several evidences support that there is a balance of pro-inflammatory and anti-inflammatory/pro-resolving pathways in normotensive pregnant women as consequence of functioning mechanisms of resolution of inflammation, leading to a state of controlled inflammatory response in these women. On the other hand, inflammation is overwhelming in PE women, probably because of dysregulated resolution of inflammation mechanisms (Fig. 3). Moreover, pro-inflammatory and anti-inflammatory/pro-resolving mediators from diverse nature might be at higher, lower, or similar levels in PE women compared with normotensive pregnant women, reinforcing the complex regulation of resolution pathways.

The apparently contradictory findings regarding the measurement of pro-resolving mediators in PE can be a mirror of the biological sample tested (serum/plasma—systemic vs. placenta—local) and moment (onset vs. established

inflammation), in which such mediators were measured. It is known that some of the pro-resolving mediators may have dual activities during the inflammatory response; i.e., they can be pro-inflammatory at the beginning of inflammation to assure proper activation of the immunologic system and, as inflammation progresses, they can be pro-resolving, acting as brakes for the inflammatory response. In addition, the activity of some mediators may be influenced by several factors, such as molecule structure (e.g., AnxA1 cleavage generates short peptides believed to have pro-inflammatory activities), the cell type in which they act (e.g., LXA4 induces apoptosis of neutrophils while rescue macrophage from death), or concentration (e.g., NO and H<sub>2</sub>S have anti-inflammatory actions at low concentrations but pro-inflammatory actions at high concentrations) [33, 113, 114, 133–135, 223, 224]. However, whether these activities may be applied in the context of PE remains to be determined.

There are few mechanistic studies in the literature for most of the pro-resolving mediators described in this work in the context of PE. Further investigation about the role of pro-resolving mediators in PE pathogenesis is warranted, for example, using knockout animals and therapeutic strategies in animal models. However, none of the available animal models of PE can mimic the full spectrum of the human disease [225]. Prospective studies with standardized methodologies would also be valuable to assess whether altered levels and/or actions of pro-resolving mediators in PE women are causes or consequences of the disease. The knowledge acquired from these studies will provide a basis for future clinical trials about novel therapies targeting pro-resolving mechanisms in PE.

**Acknowledgments** The authors are grateful to Fundação de Amparo à Pesquisa do Estado de Minas Gerais (FAPEMIG), Conselho Nacional de Desenvolvimento Científico e Tecnológico (CNPq), and Coordenação de Aperfeiçoamento de Pessoal de Nível Superior (CAPES) Research Fellowship.

## Compliance with ethical standards

**Funding** This study was funded by Fundação de Amparo à Pesquisa do Estado de Minas Gerais (FAPEMIG; APQ-03318-15), Conselho Nacional de Desenvolvimento Científico e Tecnológico (CNPq; Research Fellowship), and Coordenação de Aperfeiçoamento de Pessoal de Nível Superior (CAPES; Ph.D. scholarship).

**Conflict of interest** The authors declare that they have no conflict of interest.

## References

1. American College of Obstetricians and Gynecologists. Hypertension in pregnancy. Report of the American College of Obstetricians and Gynecologists' Task Force on Hypertension in Pregnancy. *Obstet Gynecol.* 2013;122(5):1122–31.

2. Chaiworapongsa T, Chaemsaitong P, Yeo L, Romero R. Preeclampsia part 1: current understanding of its pathophysiology. *Nat Rev Nephrol.* 2014;10(8):466–80.
3. American College of Obstetricians and Gynecologists. ACOG practice bulletin. Diagnosis and management of preeclampsia and eclampsia. Number 33, January 2002. *Obstet Gynecol.* 2002;99(1):159–67.
4. von Dadelszen P, Magee LA, Roberts JM. Subclassification of preeclampsia. *Hypertens Pregnancy.* 2003;22(2):143–8.
5. Steegers EA, von Dadelszen P, Duvekot JJ, Pijnenborg R. Preeclampsia. *Lancet.* 2010;376(9741):631–44.
6. Raymond D, Peterson E. A critical review of early-onset and late-onset preeclampsia. *Obstet Gynecol Surv.* 2011;66(8):497–506.
7. Roberts JM, Hubel CA. The two stage model of preeclampsia: variations on the theme. *Placenta.* 2009;30(Suppl A):S32–7.
8. Lam C, Lim KH, Karumanchi SA. Circulating angiogenic factors in the pathogenesis and prediction of preeclampsia. *Hypertension.* 2005;46(5):1077–85.
9. Ahmed A, Ramma W. Unravelling the theories of pre-eclampsia: are the protective pathways the new paradigm? *Br J Pharmacol.* 2015;172(6):1574–86.
10. Cotechini T, Komisarenko M, Sperou A, Macdonald-Goodfellow S, Adams MA, Graham CH. Inflammation in rat pregnancy inhibits spiral artery remodeling leading to fetal growth restriction and features of preeclampsia. *J Exp Med.* 2014;211(1):165–79.
11. Matsubara K, Higaki T, Matsubara Y, Nawa A. Nitric oxide and reactive oxygen species in the pathogenesis of preeclampsia. *Int J Mol Sci.* 2015;16(3):4600–14.
12. Wallace JL, Ianaro A, Flannigan KL, Cirino G. Gaseous mediators in resolution of inflammation. *Semin Immunol.* 2015;27(3):227–33.
13. Chovatiya R, Medzhitov R. Stress, inflammation, and defense of homeostasis. *Mol Cell.* 2014;54(2):281–8.
14. Serhan CN, Savill J. Resolution of inflammation: the beginning programs the end. *Nat Immunol.* 2005;6(12):1191–7.
15. Serhan CN, Brain SD, Buckley CD, Gilroy DW, Haslett C, O'Neill LA, et al. Resolution of inflammation: state of the art, definitions and terms. *FASEB J.* 2007;21(2):325–32.
16. Headland SE, Norling LV. The resolution of inflammation: principles and challenges. *Semin Immunol.* 2015;27(3):149–60.
17. Sugimoto M, Sousa L, Pinho V, Perretti M, Teixeira M. Resolution of inflammation: what controls its onset? *Front Immunol.* 2016;160(7):1–18.
18. Vago JP, Tavares LP, Sugimoto MA, Lima GL, Galvão I, de Caux TR, et al. Proresolving actions of synthetic and natural protease inhibitors are mediated by Annexin A1. *J Immunol.* 2016;196(4):1922–32.
19. Odaka C, Mizuochi T, Yang J, Ding A. Murine macrophages produce secretory leukocyte protease inhibitor during clearance of apoptotic cells: implications for resolution of the inflammatory response. *J Immunol.* 2003;171(3):1507–14.
20. Titos E, Rius B, González-Pérez A, López-Vicario C, Morán-Salvador E, Martínez-Clemente M, et al. Resolvin D1 and its precursor docosahexaenoic acid promote resolution of adipose tissue inflammation by eliciting macrophage polarization toward an M2-like phenotype. *J Immunol.* 2011;187(10):5408–18.
21. Fullerton JN, Gilroy DW. Resolution of inflammation: a new therapeutic frontier. *Nat Rev Drug Discov.* 2016;15(8):551–67.
22. Orsi NM. Cytokine networks in the establishment and maintenance of pregnancy. *Hum Fertil (Camb).* 2008;11(4):222–30.
23. Redman CW, Sacks GP, Sargent IL. Preeclampsia: an excessive maternal inflammatory response to pregnancy. *Am J Obstet Gynecol.* 1999;180(2 Pt 1):499–506.
24. Schminkey DL, Groer M. Imitating a stress response: a new hypothesis about the innate immune system's role in pregnancy. *Med Hypotheses.* 2014;82(6):721–9.
25. Saito S, Nakashima A, Shima T, Ito M. Th1/Th2/Th17 and regulatory T-cell paradigm in pregnancy. *Am J Reprod Immunol.* 2010;63(6):601–10.
26. Li M, Piao L, Chen CP, Wu X, Yeh CC, Masch R, et al. Modulation of decidual macrophage polarization by macrophage colony-stimulating factor derived from first-trimester decidual cells: implication in preeclampsia. *Am J Pathol.* 2016;186(5):1258–66.
27. Lee CL, Guo Y, So KH, Vijayan M, Wong VH, Yao Y, et al. Soluble human leukocyte antigen G5 polarizes differentiation of macrophages toward a decidual macrophage-like phenotype. *Hum Reprod.* 2015;30(10):2263–74.
28. Perretti M, D'Acquisto F. Annexin A1 and glucocorticoids as effectors of the resolution of inflammation. *Nat Rev Immunol.* 2009;9(1):62–70.
29. Sugimoto MA, Vago JP, Teixeira MM, Sousa LP. Annexin A1 and the resolution of inflammation: modulation of neutrophil recruitment, apoptosis, and clearance. *J Immunol Res.* 2016;2016:8239258.
30. John CD, Gavins FN, Buss NA, Cover PO, Buckingham JC. Annexin A1 and the formyl peptide receptor family: neuroendocrine and metabolic aspects. *Curr Opin Pharmacol.* 2008;8(6):765–76.
31. John CD, Christian HC, Morris JF, Flower RJ, Solito E, Buckingham JC. Annexin I and the regulation of endocrine function. *Trends Endocrinol Metab.* 2004;15(3):103–9.
32. Eke Gungor H, Tahan F, Gokahmetoglu S, Saraymen B. Decreased levels of lipoxin A4 and annexin A1 in wheezy infants. *Int Arch Allergy Immunol.* 2014;163(3):193–7.
33. Williams SL, Milne IR, Bagley CJ, Gamble JR, Vadas MA, Pitson SM, et al. A proinflammatory role for proteolytically cleaved annexin A1 in neutrophil transendothelial migration. *J Immunol.* 2010;185(5):3057–63.
34. Tsao FH, Meyer KC, Chen X, Rosenthal NS, Hu J. Degradation of annexin I in bronchoalveolar lavage fluid from patients with cystic fibrosis. *Am J Respir Cell Mol Biol.* 1998;18(1):120–8.
35. Cooray SN, Gobbetti T, Montero-Melendez T, McArthur S, Thompson D, Clark AJ, et al. Ligand-specific conformational change of the G-protein-coupled receptor ALX/FPR2 determines proresolving functional responses. *Proc Natl Acad Sci U S A.* 2013;110(45):18232–7.
36. Planagumà A, Kazani S, Marigowda G, Haworth O, Mariani TJ, Israel E, et al. Airway lipoxin A4 generation and lipoxin A4 receptor expression are decreased in severe asthma. *Am J Respir Crit Care Med.* 2008;178(6):574–82.
37. Perucci LO, Carneiro FS, Ferreira CN, Sugimoto MA, Soriani FM, Martins GG, et al. Annexin A1 is increased in the plasma of preeclamptic women. *PLoS One.* 2015;10(9):e0138475.
38. Xu Z, Zhao F, Lin F, Xiang H, Wang N, Ye D, et al. Preeclampsia is associated with a deficiency of lipoxin A4, an endogenous anti-inflammatory mediator. *Fertil Steril.* 2014;102(1):282–90.e4.
39. Dong W, Yin L. Expression of lipoxin A4, TNF $\alpha$  and IL-1 $\beta$  in maternal peripheral blood, umbilical cord blood and placenta, and their significance in pre-eclampsia. *Hypertens Pregnancy.* 2014;33(4):449–56.
40. Behrouz GF, Farzaneh GS, Leila J, Jaleh Z, Eskandar KS. Presence of auto-antibody against two placental proteins, annexin A1 and vitamin D binding protein, in sera of women with preeclampsia. *J Reprod Immunol.* 2013;99(1–2):10–6.
41. Canzoneri BJ, Lewis DF, Groome L, Wang Y. Increased neutrophil numbers account for leukocytosis in women with preeclampsia. *Am J Perinatol.* 2009;26(10):729–32.
42. Gupta AK, Gebhardt S, Hillermann R, Holzgreve W, Hahn S. Analysis of plasma elastase levels in early and late onset preeclampsia. *Arch Gynecol Obstet.* 2006;273(4):239–42.

43. Salama RH, Fathalla MM, Mekki AR, Elsadek B-K. Implication of umbilical cord in preeclampsia. *Med Princ Pract.* 2011;20(2):124–8.
44. Sun M, Liu Y, Gibb W. Distribution of annexin I and II in term human fetal membranes, decidua and placenta. *Placenta.* 1996;17(2–3):181–4.
45. Leffler H, Carlsson S, Hedlund M, Qian Y, Poirier F. Introduction to galectins. *Glycoconj J.* 2004;19(7–9):433–40.
46. Sato S, St-Pierre C, Bhaumik P, Nieminen J. Galectins in innate immunity: dual functions of host soluble beta-galactoside-binding lectins as damage-associated molecular patterns (DAMPs) and as receptors for pathogen-associated molecular patterns (PAMPs). *Immunol Rev.* 2009;230(1):172–87.
47. Rubinstein N, Ilarregui JM, Toscano MA, Rabinovich GA. The role of galectins in the initiation, amplification and resolution of the inflammatory response. *Tissue Antigens.* 2004;64(1):1–12.
48. Arikawa T, Simamura E, Shimada H, Nakamura T, Hatta T, Shoji H. Significance of sugar chain recognition by galectins and its involvement in disease-associated glycosylation. *Congenit Anom (Kyoto).* 2014;54(2):77–81.
49. Vasta GR. Galectins as pattern recognition receptors: structure, function, and evolution. *Adv Exp Med Biol.* 2012;946:21–36.
50. Romaniuk MA, Negrotto S, Campetella O, Rabinovich GA, Schattner M. Identification of galectins as novel regulators of platelet signaling and function. *IUBMB Life.* 2011;63(7):521–7.
51. Blois SM, Conrad ML, Freitag N, Barrientos G. Galectins in angiogenesis: consequences for gestation. *J Reprod Immunol.* 2015;108:33–41.
52. Blois SM, Ilarregui JM, Tometten M, Garcia M, Orsal AS, Cordo-Russo R, et al. A pivotal role for galectin-1 in fetomaternal tolerance. *Nat Med.* 2007;13(12):1450–7.
53. van der Leij J, van den Berg A, Blokzijl T, Harms G, van Goor H, Zwiers P, et al. Dimeric galectin-1 induces IL-10 production in T-lymphocytes: an important tool in the regulation of the immune response. *J Pathol.* 2004;204(5):511–8.
54. Kopcow HD, Rosetti F, Leung Y, Allan DS, Kutok JL, Strominger JL. T cell apoptosis at the maternal-fetal interface in early human pregnancy, involvement of galectin-1. *Proc Natl Acad Sci U S A.* 2008;105(47):18472–7.
55. Rostoker R, Yaseen H, Schiff-Zuck S, Lichtenstein RG, Rabinovich GA, Ariel A. Galectin-1 induces 12/15-lipoxygenase expression in murine macrophages and favors their conversion toward a pro-resolving phenotype. *Prostaglandins Other Lipid Mediat.* 2013;107:85–94.
56. Jeschke U, Mayr D, Schiessl B, Mylonas I, Schulze S, Kuhn C, et al. Expression of galectin-1, -3 (gal-1, gal-3) and the Thomsen-Friedenreich (TF) antigen in normal, IUGR, preeclamptic and HELLP placentas. *Placenta.* 2007;28(11–12):1165–73.
57. Than NG, Erez O, Wildman DE, Tarca AL, Edwin SS, Abbas A, et al. Severe preeclampsia is characterized by increased placental expression of galectin-1. *J Matern Fetal Neonatal Med.* 2008;21(7):429–42.
58. Freitag N, Tirado-González I, Barrientos G, Herse F, Thijssen VL, Weedon-Fekjær SM, et al. Interfering with Gal-1-mediated angiogenesis contributes to the pathogenesis of preeclampsia. *Proc Natl Acad Sci U S A.* 2013;110(28):11451–6.
59. Blois SM, Dechend R, Barrientos G, Staff AC. A potential pathophysiological role for galectins and the renin-angiotensin system in preeclampsia. *Cell Mol Life Sci.* 2015;72(1):39–50.
60. Molvarec A, Blois SM, Stenczer B, Toldi G, Tirado-Gonzalez I, Ito M, et al. Peripheral blood galectin-1-expressing T and natural killer cells in normal pregnancy and preeclampsia. *Clin Immunol.* 2011;139(1):48–56.
61. Than NG, Pick E, Bellyei S, Szigeti A, Burger O, Berente Z, et al. Functional analyses of placental protein 13/galectin-13. *Eur J Biochem.* 2004;271(6):1065–78.
62. Orendi K, Gauster M, Moser G, Meiri H, Huppertz B. The choriocarcinoma cell line BeWo: syncytial fusion and expression of syncytium-specific proteins. *Reproduction.* 2010;140(5):759–66.
63. Than NG, Romero R, Xu Y, Erez O, Xu Z, Bhatti G, et al. Evolutionary origins of the placental expression of chromosome 19 cluster galectins and their complex dysregulation in preeclampsia. *Placenta.* 2014;35(11):855–65.
64. Than NG, Romero R, Goodman M, Weckle A, Xing J, Dong Z, et al. A primate subfamily of galectins expressed at the maternal-fetal interface that promote immune cell death. *Proc Natl Acad Sci U S A.* 2009;106(24):9731–6.
65. Kliman HJ, Sammar M, Grimpel YI, Lynch SK, Milano KM, Pick E, et al. Placental protein 13 and decidual zones of necrosis: an immunologic diversion that may be linked to preeclampsia. *Reprod Sci.* 2012;19(1):16–30.
66. Than NG, Abdul Rahman O, Magenheimer R, Nagy B, Fule T, Hargitai B, et al. Placental protein 13 (galectin-13) has decreased placental expression but increased shedding and maternal serum concentrations in patients presenting with preterm pre-eclampsia and HELLP syndrome. *Virchows Arch.* 2008;453(4):387–400.
67. Huppertz B, Sammar M, Chefetz I, Neumaier-Wagner P, Bartz C, Meiri H. Longitudinal determination of serum placental protein 13 during development of preeclampsia. *Fetal Diagn Ther.* 2008;24(3):230–6.
68. Gonen R, Shahar R, Grimpel YI, Chefetz I, Sammar M, Meiri H, et al. Placental protein 13 as an early marker for pre-eclampsia: a prospective longitudinal study. *BJOG.* 2008;115(12):1465–72.
69. Sekizawa A, Purwosunu Y, Yoshimura S, Nakamura M, Shimizu H, Okai T, et al. PP13 mRNA expression in trophoblasts from preeclamptic placentas. *Reprod Sci.* 2009;16(4):408–13.
70. Shimizu H, Sekizawa A, Purwosunu Y, Nakamura M, Farina A, Rizzo N, et al. PP13 mRNA expression in the cellular component of maternal blood as a marker for preeclampsia. *Prenat Diagn.* 2009;29(13):1231–6.
71. Farina A, Zucchini C, Sekizawa A, Purwosunu Y, de Sanctis P, Santarsiero G, et al. Performance of messenger RNAs circulating in maternal blood in the prediction of preeclampsia at 10–14 weeks. *Am J Obstet Gynecol.* 2010;203(6):575.e1–7.
72. Gebhardt S, Bruiners N, Hillermann R. A novel exonic variant (221delT) in the LGALS13 gene encoding placental protein 13 (PP13) is associated with preterm labour in a low risk population. *J Reprod Immunol.* 2009;82(2):166–73.
73. Than NG, Balogh A, Romero R, Kárpáti E, Erez O, Szilágyi A, et al. Placental protein 13 (PP13)—a placental immunoregulatory galectin protecting pregnancy. *Front Immunol.* 2014;5:348.
74. Huppertz B, Meiri H, Gizurarson S, Osol G, Sammar M. Placental protein 13 (PP13): a new biological target shifting individualized risk assessment to personalized drug design combating preeclampsia. *Hum Reprod Update.* 2013;19(4):391–405.
75. Sammar M, Nisemblat S, Fleischfarb Z, Golan A, Sadan O, Meiri H, et al. Placenta-bound and body fluid PP13 and its mRNA in normal pregnancy compared to preeclampsia, HELLP and preterm delivery. *Placenta.* 2011;32(Suppl):S30–6.
76. Zabel BA, Kwitniewski M, Banas M, Zabieglo K, Murzyn K, Cichy J. Chemerin regulation and role in host defense. *Am J Clin Exp Immunol.* 2014;3(1):1–19.
77. Carlino C, Trotta E, Stabile H, Morrone S, Bulla R, Soriani A, et al. Chemerin regulates NK cell accumulation and endothelial cell morphogenesis in the decidua during early pregnancy. *J Clin Endocrinol Metab.* 2012;97(10):3603–12.
78. Tessier DR, Yockell-Lelievre J, Gruslin A. Uterine spiral artery remodeling: the role of uterine natural killer cells and extravillous trophoblasts in normal and high-risk human pregnancies. *Am J Reprod Immunol.* 2015;74(1):1–11.
79. Kaur J, Adya R, Tan BK, Chen J, Randeve HS. Identification of chemerin receptor (ChemR23) in human endothelial cells:



- chemerin-induced endothelial angiogenesis. *Biochem Biophys Res Commun.* 2010;391(4):1762–8.
80. Mariani F, Roncucci L. Chemerin/chemR23 axis in inflammation onset and resolution. *Inflamm Res.* 2015;64(2):85–95.
  81. Moretta A, Marcenaro E, Parolini S, Ferlazzo G, Moretta L. NK cells at the interface between innate and adaptive immunity. *Cell Death Differ.* 2008;15(2):226–33.
  82. Hespel C, Moser M. Role of inflammatory dendritic cells in innate and adaptive immunity. *Eur J Immunol.* 2012;42(10):2535–43.
  83. Zabel BA, Allen SJ, Kulig P, Allen JA, Cichy J, Handel TM, et al. Chemerin activation by serine proteases of the coagulation, fibrinolytic, and inflammatory cascades. *J Biol Chem.* 2005;280(41):34661–6.
  84. Garces MF, Sanchez E, Ruiz-Parra AI, Rubio-Romero JA, Angel-Muller E, Suarez MA, et al. Serum chemerin levels during normal human pregnancy. *Peptides.* 2013;42:138–43.
  85. Kasher-Meron M, Mazaki-Tovi S, Barhod E, Hemi R, Haas J, Gat I, et al. Chemerin concentrations in maternal and fetal compartments: implications for metabolic adaptations to normal human pregnancy. *J Perinat Med.* 2014;42(3):371–8.
  86. Wang L, Yang T, Ding Y, Zhong Y, Yu L, Peng M. Chemerin plays a protective role by regulating human umbilical vein endothelial cell-induced nitric oxide signaling in preeclampsia. *Endocrine.* 2015;48(1):299–308.
  87. Duan DM, Niu JM, Lei Q, Lin XH, Chen X. Serum levels of the adipokine chemerin in preeclampsia. *J Perinat Med.* 2012;40(2):121–7.
  88. Stepan H, Philipp A, Roth I, Kralisch S, Jank A, Schaarschmidt W, et al. Serum levels of the adipokine chemerin are increased in preeclampsia during and 6 months after pregnancy. *Regul Pept.* 2011;168(1–3):69–72.
  89. Xu QL, Zhu M, Jin Y, Wang N, Xu HX, Quan LM, et al. The predictive value of the first-trimester maternal serum chemerin level for pre-eclampsia. *Peptides.* 2014;62:150–4.
  90. Serhan CN. Pro-resolving lipid mediators are leads for resolution physiology. *Nature.* 2014;510(7503):92–101.
  91. Claria J, Serhan CN. Aspirin triggers previously undescribed bioactive eicosanoids by human endothelial cell-leukocyte interactions. *Proc Natl Acad Sci U S A.* 1995;92(21):9475–9.
  92. Serhan CN. Lipoxins and aspirin-triggered 15-epi-lipoxins are the first lipid mediators of endogenous anti-inflammation and resolution. *Prostaglandins Leukot Essent Fatty Acids.* 2005;73(3–4):141–62.
  93. Fierro IM. Angiogenesis and lipoxins. *Prostaglandins Leukot Essent Fatty Acids.* 2005;73(3–4):271–5.
  94. Choi G, Hwang SW. Modulation of the activities of neuronal ion channels by fatty acid-derived pro-resolvents. *Front Physiol.* 2016;7:523.
  95. Levy BD, Serhan CN. Exploring new approaches to the treatment of asthma: potential roles for lipoxins and aspirin-triggered lipid mediators. *Drugs Today (Barc).* 2003;39(5):373–84.
  96. Lee TH. Lipoxin A4: a novel anti-inflammatory molecule? *Thorax.* 1995;50(2):111–2.
  97. Gavins FN, Sawmynaden P, Chatterjee BE, Perretti M. A twist in anti-inflammation: annexin 1 acts via the lipoxin A4 receptor. *Prostaglandins Leukot Essent Fatty Acids.* 2005;73(3–4):211–9.
  98. Wang J, Huang Y, Zhou J, Liu X. Effect of lipoxin A<sub>4</sub> on IL-1 $\beta$  production of monocytes and its possible mechanism in severe preeclampsia. *J Huazhong Univ Sci Technolog Med Sci.* 2010;30(6):767–70.
  99. Gil-Villa AM, Norling LV, Serhan CN, Cordero D, Rojas M, Cadavid A. Aspirin triggered-lipoxin A4 reduces the adhesion of human polymorphonuclear neutrophils to endothelial cells initiated by preeclamptic plasma. *Prostaglandins Leukot Essent Fatty Acids.* 2012;87(4–5):127–34.
  100. Lin F, Zeng P, Xu Z, Ye D, Yu X, Wang N, et al. Treatment of lipoxin a(4) and its analogue on low-dose endotoxin induced preeclampsia in rat and possible mechanisms. *Reprod Toxicol.* 2012;34(4):677–85.
  101. Huang LL, Su S, Awale R, Zhang XY, Zhong LL, Tang H. Expression of anti-inflammatory mediator lipoxin A4 and inflammation responsive transcription factors NF-kappa B in patients with preeclampsia. *Clin Exp Obstet Gynecol.* 2014;41(5):561–6.
  102. Perucci LO, Santos PC, Ribeiro LS, Souza DG, Gomes KB, Dusse LM, et al. Lipoxin A4 is increased in the plasma of preeclamptic women. *Am J Hypertens.* 2016;29(10):1179–85.
  103. Cooper CE, Brown GC. The inhibition of mitochondrial cytochrome oxidase by the gases carbon monoxide, nitric oxide, hydrogen cyanide and hydrogen sulfide: chemical mechanism and physiological significance. *J Bioenerg Biomembr.* 2008;40(5):533–9.
  104. Wang R. Two's company, three's a crowd: can H2S be the third endogenous gaseous transmitter? *FASEB J.* 2002;16(13):1792–8.
  105. Kajimura M, Fukuda R, Bateman RM, Yamamoto T, Suematsu M. Interactions of multiple gas-transducing systems: hallmarks and uncertainties of CO, NO, and H<sub>2</sub>S gas biology. *Antioxid Redox Signal.* 2010;13(2):157–92.
  106. Lyall F. Development of the utero-placental circulation: the role of carbon monoxide and nitric oxide in trophoblast invasion and spiral artery transformation. *Microsc Res Tech.* 2003;60(4):402–11.
  107. Li H, Horke S, Förstermann U. Vascular oxidative stress, nitric oxide and atherosclerosis. *Atherosclerosis.* 2014;237(1):208–19.
  108. Nagy G, Koncz A, Telarico T, Fernandez D, Ersek B, Buzás E, et al. Central role of nitric oxide in the pathogenesis of rheumatoid arthritis and systemic lupus erythematosus. *Arthritis Res Ther.* 2010;12(3):210.
  109. Ignarro LJ, Buga GM, Wood KS, Byrns RE, Chaudhuri G. Endothelium-derived relaxing factor produced and released from artery and vein is nitric oxide. *Proc Natl Acad Sci U S A.* 1987;84(24):9265–9.
  110. Sogo N, Magid KS, Shaw CA, Webb DJ, Megson IL. Inhibition of human platelet aggregation by nitric oxide donor drugs: relative contribution of cGMP-independent mechanisms. *Biochem Biophys Res Commun.* 2000;279(2):412–9.
  111. Ward C, Wong TH, Murray J, Rahman I, Haslett C, Chilvers ER, et al. Induction of human neutrophil apoptosis by nitric oxide donors: evidence for a caspase-dependent, cyclic-GMP-independent, mechanism. *Biochem Pharmacol.* 2000;59(3):305–14.
  112. Lo Faro ML, Fox B, Whatmore JL, Winyard PG, Whiteman M. Hydrogen sulfide and nitric oxide interactions in inflammation. *Nitric Oxide.* 2014;41:38–47.
  113. Laroux FS, Lefer DJ, Kawachi S, Scalia R, Cockrell AS, Gray L, et al. Role of nitric oxide in the regulation of acute and chronic inflammation. *Antioxid Redox Signal.* 2000;2(3):391–6.
  114. Guzik TJ, Korb R, Adamek-Guzik T. Nitric oxide and superoxide in inflammation and immune regulation. *J Physiol Pharmacol.* 2003;54(4):469–87.
  115. Shaw CA, Taylor EL, Fox S, Megson IL, Rossi AG. Differential susceptibility to nitric oxide-evoked apoptosis in human inflammatory cells. *Free Radic Biol Med.* 2011;50(1):93–101.
  116. Huang LT, Hsieh CS, Chang KA, Tain YL. Roles of nitric oxide and asymmetric dimethylarginine in pregnancy and fetal programming. *Int J Mol Sci.* 2012;13(11):14606–22.
  117. Morris NH, Sooranna SR, Lammont JG, Poston L, Ramsey B, Pearson JD, et al. Nitric oxide synthase activities in placental tissue from normotensive, pre-eclamptic and growth retarded pregnancies. *Br J Obstet Gynaecol.* 1995;102(9):711–4.
  118. Böger RH, Diemert A, Schwedhelm E, Lüneburg N, Maas R, Hecher K. The role of nitric oxide synthase inhibition by

- asymmetric dimethylarginine in the pathophysiology of preeclampsia. *Gynecol Obstet Investig.* 2010;69(1):1–13.
119. López-Jaramillo P, Arenas WD, García RG, Rincon MY, López M. The role of the L-arginine-nitric oxide pathway in preeclampsia. *Ther Adv Cardiovasc Dis.* 2008;2(4):261–75.
  120. Bian Z, Shixia C, Duan T. First-trimester maternal serum levels of sFLT1, PGF and ADMA predict preeclampsia. *PLoS One.* 2015;10(4):e0124684.
  121. Alpoim PN, Godoi LC, Freitas LG, Gomes KB, Dusse LM. Assessment of L-arginine asymmetric 1 dimethyl (ADMA) in early-onset and late-onset (severe) preeclampsia. *Nitric Oxide.* 2013;33:81–2.
  122. Alpoim PN, Gomes KB, Pinheiro MB, Godoi LC, Jardim LL, Muniz LG, et al. Polymorphisms in endothelial nitric oxide synthase gene in early and late severe preeclampsia. *Nitric Oxide.* 2014;42:19–23.
  123. Doridot L, Passet B, Mehats C, Rigourd V, Barbaux S, Ducat A, et al. Preeclampsia-like symptoms induced in mice by fetoplacental expression of STOX1 are reversed by aspirin treatment. *Hypertension.* 2013;61(3):662–8.
  124. van Dijk M, Mulders J, Poutsma A, Konst AA, Lachmeijer AM, Dekker GA, et al. Maternal segregation of the Dutch preeclampsia locus at 10q22 with a new member of the winged helix gene family. *Nat Genet.* 2005;37(5):514–9.
  125. Doridot L, Chatre L, Ducat A, Vilotte JL, Lombes A, Mehats C, et al. Nitroso-redox balance and mitochondrial homeostasis are regulated by STOX1, a pre-eclampsia-associated gene. *Antioxid Redox Signal.* 2014;21(6):819–34.
  126. van Dijk M, van Bezu J, van Abel D, Dunk C, Blankenstein MA, Oudejans CB, et al. The STOX1 genotype associated with preeclampsia leads to a reduction of trophoblast invasion by alpha-T-catenin upregulation. *Hum Mol Genet.* 2010;19(13):2658–67.
  127. Nanaev A, Chwalisz K, Frank HG, Kohnen G, Hegele-Hartung C, Kaufmann P. Physiological dilation of uteroplacental arteries in the guinea pig depends on nitric oxide synthase activity of extravillous trophoblast. *Cell Tissue Res.* 1995;282(3):407–21.
  128. Holwerda KM, Faas MM, van Goor H, Lely AT. Gasotransmitters: a solution for the therapeutic dilemma in preeclampsia? *Hypertension.* 2013;62(4):653–9.
  129. Meher S, Duley L. Nitric oxide for preventing pre-eclampsia and its complications. *Cochrane Database Syst Rev.* 2007;2:CD006490.
  130. Olas B. Hydrogen sulfide in signaling pathways. *Clin Chim Acta.* 2015;439:212–8.
  131. Kida M, Sugiyama T, Yoshimoto T, Ogawa Y. Hydrogen sulfide increases nitric oxide production with calcium-dependent activation of endothelial nitric oxide synthase in endothelial cells. *Eur J Pharm Sci.* 2013;48(1–2):211–5.
  132. Szabó C. Hydrogen sulphide and its therapeutic potential. *Nat Rev Drug Discov.* 2007;6(11):917–35.
  133. Collin M, Anuar FB, Murch O, Bhatia M, Moore PK, Thiemermann C. Inhibition of endogenous hydrogen sulfide formation reduces the organ injury caused by endotoxemia. *Br J Pharmacol.* 2005;146(4):498–505.
  134. Zhang H, Zhi L, Moochhala S, Moore PK, Bhatia M. Hydrogen sulfide acts as an inflammatory mediator in cecal ligation and puncture-induced sepsis in mice by upregulating the production of cytokines and chemokines via NF-kappaB. *Am J Physiol Lung Cell Mol Physiol.* 2007;292(4):L960–71.
  135. Wallace JL, Vong L, McKnight W, Dickey M, Martin GR. Endogenous and exogenous hydrogen sulfide promotes resolution of colitis in rats. *Gastroenterology.* 2009;137(2):569–78. **78.e1**
  136. Mariggio MA, Minunno V, Riccardi S, Santacroce R, De Rinaldis P, Fumarulo R. Sulfide enhancement of PMN apoptosis. *Immunopharmacol Immunotoxicol.* 1998;20(3):399–408.
  137. Miao L, Shen X, Whiteman M, Xin H, Shen Y, Xin X, et al. Hydrogen sulfide mitigates myocardial infarction via promotion of mitochondrial biogenesis-dependent M2 polarization of macrophages. *Antioxid Redox Signal.* 2016;25(5):268–81.
  138. Dufton N, Natividad J, Verdu EF, Wallace JL. Hydrogen sulfide and resolution of acute inflammation: a comparative study utilizing a novel fluorescent probe. *Sci Rep.* 2012;2:499.
  139. Brancaleone V, Mitidieri E, Flower RJ, Cirino G, Perretti M. Annexin A1 mediates hydrogen sulfide properties in the control of inflammation. *J Pharmacol Exp Ther.* 2014;351(1):96–104.
  140. Papapetropoulos A, Pyriochou A, Altaany Z, Yang G, Marazioti A, Zhou Z, et al. Hydrogen sulfide is an endogenous stimulator of angiogenesis. *Proc Natl Acad Sci U S A.* 2009;106(51):21972–7.
  141. Patel P, Vatish M, Heptinstall J, Wang R, Carson RJ. The endogenous production of hydrogen sulphide in intrauterine tissues. *Reprod Biol Endocrinol.* 2009;7:10.
  142. Wang K, Ahmad S, Cai M, Rennie J, Fujisawa T, Crispi F, et al. Dysregulation of hydrogen sulfide producing enzyme cystathionine  $\gamma$ -lyase contributes to maternal hypertension and placental abnormalities in preeclampsia. *Circulation.* 2013;127(25):2514–22.
  143. Cindrova-Davies T, Herrera EA, Niu Y, Kingdom J, Giussani DA, Burton GJ. Reduced cystathionine  $\gamma$ -lyase and increased miR-21 expression are associated with increased vascular resistance in growth-restricted pregnancies: hydrogen sulfide as a placental vasodilator. *Am J Pathol.* 2013;182(4):1448–58.
  144. Holwerda KM, Bos EM, Rajakumar A, Ris-Stalpers C, van Pampus MG, Timmer A, et al. Hydrogen sulfide producing enzymes in pregnancy and preeclampsia. *Placenta.* 2012;33(6):518–21.
  145. Roes EM, Raijmakers MT, Boo TM, Zusterzeel PL, Merkus HM, Peters WH, et al. Oral N-acetylcysteine administration does not stabilise the process of established severe preeclampsia. *Eur J Obstet Gynecol Reprod Biol.* 2006;127(1):61–7.
  146. Ryter SW, Choi AM. Heme oxygenase-1/carbon monoxide: from metabolism to molecular therapy. *Am J Respir Cell Mol Biol.* 2009;41(3):251–60.
  147. Wu L, Wang R. Carbon monoxide: endogenous production, physiological functions, and pharmacological applications. *Pharmacol Rev.* 2005;57(4):585–630.
  148. Bilban M, Haschemi A, Wegiel B, Chin BY, Wagner O, Otterbein LE. Heme oxygenase and carbon monoxide initiate homeostatic signaling. *J Mol Med (Berl).* 2008;86(3):267–79.
  149. Peers C, Boyle JP, Scragg JL, Dallas ML, Al-Owais MM, Hettiarachichi NT, et al. Diverse mechanisms underlying the regulation of ion channels by carbon monoxide. *Br J Pharmacol.* 2015;172(6):1546–56.
  150. Urquhart P, Rosignoli G, Cooper D, Motterlini R, Perretti M. Carbon monoxide-releasing molecules modulate leukocyte-endothelial interactions under flow. *J Pharmacol Exp Ther.* 2007;321(2):656–62.
  151. Morse D, Pischke SE, Zhou Z, Davis RJ, Flavell RA, Loop T, et al. Suppression of inflammatory cytokine production by carbon monoxide involves the JNK pathway and AP-1. *J Biol Chem.* 2003;278(39):36993–8.
  152. Otterbein LE, May A, Chin BY. Carbon monoxide increases macrophage bacterial clearance through toll-like receptor (TLR)4 expression. *Cell Mol Biol (Noisy-le-grand).* 2005;51(5):433–40.
  153. Chiang N, Shinohara M, Dalli J, Mirakaj V, Kibi M, Choi AM, et al. Inhaled carbon monoxide accelerates resolution of inflammation via unique proresolving mediator-heme oxygenase-1 circuits. *J Immunol.* 2013;190(12):6378–88.
  154. Lyall F, Barber A, Myatt L, Bulmer JN, Robson SC. Hemeoxygenase expression in human placenta and placental

- bed implies a role in regulation of trophoblast invasion and placental function. *FASEB J*. 2000;14(1):208–19.
155. Bainbridge SA, Smith GN. HO in pregnancy. *Free Radic Biol Med*. 2005;38(8):979–88.
  156. McCaig D, Lyall F. Inhibitors of heme oxygenase reduce invasion of human primary cytotrophoblast cells in vitro. *Placenta*. 2009;30(6):536–8.
  157. Sollwedel A, Bertoja AZ, Zenclussen ML, Gerlof K, Lisewski U, Wafula P, et al. Protection from abortion by heme oxygenase-1 up-regulation is associated with increased levels of bag-1 and neuropilin-1 at the fetal-maternal interface. *J Immunol*. 2005;175(8):4875–85.
  158. Baum M, Schiff E, Kreiser D, Dennery PA, Stevenson DK, Rosenthal T, et al. End-tidal carbon monoxide measurements in women with pregnancy-induced hypertension and preeclampsia. *Am J Obstet Gynecol*. 2000;183(4):900–3.
  159. Yusuf K, Kamaluddeen M, Wilson RD, Akierman A. Carboxyhemoglobin levels in umbilical cord blood of women with pre-eclampsia and intrauterine growth restriction. *J Perinat Med*. 2012;40(6):619–24.
  160. Zhai D, Guo Y, Smith G, Krewski D, Walker M, Wen SW. Maternal exposure to moderate ambient carbon monoxide is associated with decreased risk of preeclampsia. *Am J Obstet Gynecol*. 2012;207(1):57.e1–9.
  161. Wikström AK, Stephansson O, Cnattingius S. Tobacco use during pregnancy and preeclampsia risk: effects of cigarette smoking and snuff. *Hypertension*. 2010;55(5):1254–9.
  162. Maebayashi Asanuma A, Yamamoto T, Azuma H, Kato E, Yamamoto N, Murase T, et al. Expression of placenta growth factor, soluble fms-like tyrosine kinase-1, metal-responsive transcription factor-1, heme oxygenase 1 and hypoxia inducible factor-1 $\alpha$  mRNAs in pre-eclampsia placenta and the effect of pre-eclampsia sera on their expression of choriocarcinoma cells. *J Obstet Gynaecol Res*. 2014;40(10):2095–103.
  163. Zenclussen AC, Lim E, Knoeller S, Knackstedt M, Hertwig K, Hagen E, et al. Heme oxygenases in pregnancy II: HO-2 is down-regulated in human pathologic pregnancies. *Am J Reprod Immunol*. 2003;50(1):66–76.
  164. Barber A, Robson SC, Myatt L, Bulmer JN, Lyall F. Heme oxygenase expression in human placenta and placental bed: reduced expression of placenta endothelial HO-2 in preeclampsia and fetal growth restriction. *FASEB J*. 2001;15(7):1158–68.
  165. Eide IP, Isaksen CV, Salvesen KA, Langaas M, Schönberg SA, Austgulen R. Decidual expression and maternal serum levels of heme oxygenase 1 are increased in pre-eclampsia. *Acta Obstet Gynecol Scand*. 2008;87(3):272–9.
  166. Tong S, Kaitu'u-Lino TJ, Onda K, Beard S, Hastie R, Binder NK, et al. Heme oxygenase-1 is not decreased in preeclamptic placenta and does not negatively regulate placental soluble fms-like tyrosine kinase-1 or soluble endoglin secretion. *Hypertension*. 2015;66(5):1073–81.
  167. Brownfoot FC, Tong S, Hannan NJ, Binder NK, Walker SP, Cannon P, et al. Effects of pravastatin on human placenta, endothelium, and women with severe preeclampsia. *Hypertension*. 2015;66(3):687–97. **discussion 445**
  168. Ramma W, Ahmed A. Therapeutic potential of statins and the induction of heme oxygenase-1 in preeclampsia. *J Reprod Immunol*. 2014;101-102:153–60.
  169. McCarthy FP, Drewlo S, Kingdom J, Johns EJ, Walsh SK, Kenny LC. Peroxisome proliferator-activated receptor- $\gamma$  as a potential therapeutic target in the treatment of preeclampsia. *Hypertension*. 2011;58(2):280–6.
  170. Venditti CC, Casselman R, Young I, Karumanchi SA, Smith GN. Carbon monoxide prevents hypertension and proteinuria in an adenovirus sFlt-1 preeclampsia-like mouse model. *PLoS One*. 2014;9(9):e106502.
  171. Picciotto MR, Higley MJ, Mineur YS. Acetylcholine as a neuromodulator: cholinergic signaling shapes nervous system function and behavior. *Neuron*. 2012;76(1):116–29.
  172. Prado MA, Reis RA, Prado VF, de Mello MC, Gomez MV, de Mello FG. Regulation of acetylcholine synthesis and storage. *Neurochem Int*. 2002;41(5):291–9.
  173. Kellogg Jr DL, Zhao JL, Coey U, Green JV. Acetylcholine-induced vasodilation is mediated by nitric oxide and prostaglandins in human skin. *J Appl Physiol* (1985). 2005;98(2):629–32.
  174. Laurent P, Safar ME, Meaune S, Blacher J. Influence of L-nitroarginine methyl ester, acetylcholine, and adenosine on mean blood pressure, pulse pressure, and pulse pressure amplification in rats. *J Cardiovasc Pharmacol*. 2003;41(2):210–8.
  175. Andersson U, Tracey KJ. Reflex principles of immunological homeostasis. *Annu Rev Immunol*. 2012;30:313–35.
  176. Andersson U, Tracey KJ. Neural reflexes in inflammation and immunity. *J Exp Med*. 2012;209(6):1057–68.
  177. Sundman E, Olofsson PS. Neural control of the immune system. *Adv Physiol Educ*. 2014;38(2):135–9.
  178. Báez-Pagán CA, Delgado-Vélez M, Lasalde-Dominicci JA. Activation of the macrophage  $\alpha 7$  nicotinic acetylcholine receptor and control of inflammation. *J NeuroImmune Pharmacol*. 2015;10(3):468–76.
  179. Tsoyi K, Jang HJ, Kim JW, Chang HK, Lee YS, Pae HO, et al. Stimulation of alpha7 nicotinic acetylcholine receptor by nicotine attenuates inflammatory response in macrophages and improves survival in experimental model of sepsis through heme oxygenase-1 induction. *Antioxid Redox Signal*. 2011;14(11):2057–70.
  180. Maldifassi MC, Atienza G, Arnalich F, López-Collazo E, Cedillo JL, Martín-Sánchez C, et al. A new IRAK-M-mediated mechanism implicated in the anti-inflammatory effect of nicotine via  $\alpha 7$  nicotinic receptors in human macrophages. *PLoS One*. 2014;9(9):e108397.
  181. Huston JM, Rosas-Ballina M, Xue X, Dowling O, Ochani K, Ochani M, et al. Cholinergic neural signals to the spleen down-regulate leukocyte trafficking via CD11b. *J Immunol*. 2009;183(1):552–9.
  182. Marigiò MA, Guida L, Laforgia A, Santacroce R, Curci E, Montemurro P, et al. Nicotine effects on polymorphonuclear cell apoptosis and lipopolysaccharide-induced monocyte functions. A possible role in periodontal disease? *J Periodontol Res*. 2001;36(1):32–9.
  183. van der Zanden EP, Snoek SA, Heinsbroek SE, Stanisor OI, Verseijden C, Boeckxstaens GE, et al. Vagus nerve activity augments intestinal macrophage phagocytosis via nicotinic acetylcholine receptor alpha4beta2. *Gastroenterology*. 2009;137(3):1029–39. **39.e1-4**
  184. Lee RH, Vazquez G. Evidence for a prosurvival role of alpha-7 nicotinic acetylcholine receptor in alternatively (M2)-activated macrophages. *Physiol Rep*. 2013;1(7):e00189.
  185. Tracey KJ. Physiology and immunology of the cholinergic antiinflammatory pathway. *J Clin Invest*. 2007;117(2):289–96.
  186. Mirakaj V, Dalli J, Granja T, Rosenberger P, Serhan CN. Vagus nerve controls resolution and pro-resolving mediators of inflammation. *J Exp Med*. 2014;211(6):1037–48.
  187. Yang CC, Chao TC, Kuo TB, Yin CS, Chen HI. Preeclamptic pregnancy is associated with increased sympathetic and decreased parasympathetic control of HR. *Am J Physiol Heart Circ Physiol*. 2000;278(4):H1269–73.
  188. Dowling O, Rochelson B, Way K, Al-Abed Y, Metz CN. Nicotine inhibits cytokine production by placenta cells via NFkappaB: potential role in pregnancy-induced hypertension. *Mol Med*. 2007;13(11–12):576–83.
  189. Liu Y, Yang J, Bao J, Li X, Ye A, Zhang G, et al. Activation of the cholinergic anti-inflammatory pathway by nicotine ameliorates



- lipopolysaccharide-induced preeclampsia-like symptoms in pregnant rats. *Placenta*. 2017;49:23–32.
190. Mimura K, Tomimatsu T, Sharentuya N, Tskitishvili E, Kinugasa-Taniguchi Y, Kanagawa T, et al. Nicotine restores endothelial dysfunction caused by excess sFlt1 and sEng in an in vitro model of preeclamptic vascular endothelium: a possible therapeutic role of nicotinic acetylcholine receptor (nAChR) agonists for preeclampsia. *Am J Obstet Gynecol*. 2010;202(5):464 e1–6.
  191. Machaalani R, Ghazavi E, David RV, Hinton T, Makris A, Hennessy A. Nicotinic acetylcholine receptors (nAChR) are increased in the pre-eclamptic placenta. *Hypertens Pregnancy*. 2015;34(2):227–40.
  192. Kwon JY, Kim YH, Kim SH, Kang MH, Maeng YS, Lee KY, et al. Difference in the expression of alpha 7 nicotinic receptors in the placenta in normal versus severe preeclampsia pregnancies. *Eur J Obstet Gynecol Reprod Biol*. 2007;132(1):35–9.
  193. Bradford D, Cole SJ, Cooper HM. Netrin-1: diversity in development. *Int J Biochem Cell Biol*. 2009;41(3):487–93.
  194. Ly NP, Komatsuzaki K, Fraser IP, Tseng AA, Prodhan P, Moore KJ, et al. Netrin-1 inhibits leukocyte migration in vitro and in vivo. *Proc Natl Acad Sci U S A*. 2005;102(41):14729–34.
  195. Aherne CM, Collins CB, Masterson JC, Tizzano M, Boyle TA, Westrich JA, et al. Neuronal guidance molecule netrin-1 attenuates inflammatory cell trafficking during acute experimental colitis. *Gut*. 2012;61(5):695–705.
  196. Ranganathan PV, Jayakumar C, Mohamed R, Dong Z, Ramesh G. Netrin-1 regulates the inflammatory response of neutrophils and macrophages, and suppresses ischemic acute kidney injury by inhibiting COX-2-mediated PGE2 production. *Kidney Int*. 2013;83(6):1087–98.
  197. Tadagavadi RK, Wang W, Ramesh G. Netrin-1 regulates Th1/Th2/Th17 cytokine production and inflammation through UNC5B receptor and protects kidney against ischemia-reperfusion injury. *J Immunol*. 2010;185(6):3750–8.
  198. Ranganathan PV, Jayakumar C, Ramesh G. Netrin-1-treated macrophages protect the kidney against ischemia-reperfusion injury and suppress inflammation by inducing M2 polarization. *Am J Physiol Renal Physiol*. 2013;304(7):F948–57.
  199. Schlegel M, Köhler D, Körner A, Granja T, Straub A, Giera M, et al. The neuroimmune guidance cue netrin-1 controls resolution programs and promotes liver regeneration. *Hepatology*. 2015.
  200. Carney EF. Diabetic nephropathy: netrin-1 expression in proximal tubular epithelial cells protects against kidney inflammation and injury. *Nat Rev Nephrol*. 2012;8(12):681.
  201. Mirakaj V, Gatidou D, Pöttsch C, König K, Rosenberger P. Netrin-1 signaling dampens inflammatory peritonitis. *J Immunol*. 2011;186(1):549–55.
  202. Podjaski C, Alvarez JI, Bourbonniere L, Larouche S, Terouz S, Bin JM, et al. Netrin 1 regulates blood-brain barrier function and neuroinflammation. *Brain*. 2015;138(Pt 6):1598–612.
  203. Yang Y, Zou L, Xu KS. Expression of netrin-1 in placenta from patients with pre-eclampsia and the relation to placental angiogenesis. *Zhonghua Fu Chan Ke Za Zhi*. 2006;41(9):597–600.
  204. López-Novoa JM, Bernabeu C. The physiological role of endoglin in the cardiovascular system. *Am J Physiol Heart Circ Physiol*. 2010;299(4):H959–74.
  205. Powers JC, Odake S, Oleksyszyn J, Hori H, Ueda T, Boduszek B, et al. Proteases—structures, mechanism and inhibitors. *Agents Actions Suppl*. 1993;42:3–18.
  206. Mancek-Keber M. Inflammation-mediating proteases: structure, function in (patho) physiology and inhibition. *Protein Pept Lett*. 2014;21(12):1209–29.
  207. Greene CM, McElvaney NG. Proteases and antiproteases in chronic neutrophilic lung disease—relevance to drug discovery. *Br J Pharmacol*. 2009;158(4):1048–58.
  208. Apte SS, Parks WC. Metalloproteinases: a parade of functions in matrix biology and an outlook for the future. *Matrix Biol*. 2015;44–46:1–6.
  209. Cohen M, Meisser A, Bischof P. Metalloproteinases and human placental invasiveness. *Placenta*. 2006;27(8):783–93.
  210. Murphy G. Tissue inhibitors of metalloproteinases. *Genome Biol*. 2011;12(11):233.
  211. Khokha R, Murthy A, Weiss A. Metalloproteinases and their natural inhibitors in inflammation and immunity. *Nat Rev Immunol*. 2013;13(9):649–65.
  212. Nissinen L, Kähäri VM. Matrix metalloproteinases in inflammation. *Biochim Biophys Acta*. 2014;1840(8):2571–80.
  213. Black RA, Doedens JR, Mahimkar R, Johnson R, Guo L, Wallace A, et al. Substrate specificity and inducibility of TACE (tumour necrosis factor alpha-converting enzyme) revisited: the ala-Val preference, and induced intrinsic activity. *Biochem Soc Symp*. 2003;70:39–52.
  214. Tang J, Zarbock A, Gomez I, Wilson CL, Lefort CT, Stadtmann A, et al. Adam17-dependent shedding limits early neutrophil influx but does not alter early monocyte recruitment to inflammatory sites. *Blood*. 2011;118(3):786–94.
  215. Wang Y, Robertson JD, Walcheck B. Different signaling pathways stimulate a disintegrin and metalloprotease-17 (ADAM17) in neutrophils during apoptosis and activation. *J Biol Chem*. 2011;286(45):38980–8.
  216. Lakatos G, Hritz I, Varga MZ, Juhász M, Miheller P, Cierny G, et al. The impact of matrix metalloproteinases and their tissue inhibitors in inflammatory bowel diseases. *Dig Dis*. 2012;30(3):289–95.
  217. Ma R, Gu Y, Groome LJ, Wang Y. ADAM17 regulates TNF $\alpha$  production by placental trophoblasts. *Placenta*. 2011;32(12):975–80.
  218. Ma R, Gu B, Gu Y, Groome LJ, Wang Y. Down-regulation of TIMP3 leads to increase in TACE expression and TNF $\alpha$  production by placental trophoblast cells. *Am J Reprod Immunol*. 2014;71(5):427–33.
  219. Perucci LO, Gomes KB, Freitas LG, Godoi LC, Alpoim PN, Pinheiro MB, et al. Soluble endoglin, transforming growth factor-Beta 1 and soluble tumor necrosis factor alpha receptors in different clinical manifestations of preeclampsia. *PLoS One*. 2014;9(5):e97632.
  220. Pinheiro MB, Martins-Filho OA, Mota AP, Alpoim PN, Godoi LC, Silveira AC, et al. Severe preeclampsia goes along with a cytokine network disturbance towards a systemic inflammatory state. *Cytokine*. 2013;62(1):165–73.
  221. Vince GS, Starkey PM, Austgulen R, Kwiatkowski D, Redman CW. Interleukin-6, tumour necrosis factor and soluble tumour necrosis factor receptors in women with pre-eclampsia. *Br J Obstet Gynaecol*. 1995;102(1):20–5.
  222. Palei AC, Granger JP, Tanus-Santos JE. Matrix metalloproteinases as drug targets in preeclampsia. *Curr Drug Targets*. 2013;14(3):325–34.
  223. Prieto P, Cuenca J, Traves PG, Fernandez-Velasco M, Martin-Sanz P, Bosca L. Lipoxin A4 impairment of apoptotic signaling in macrophages: implication of the PI3K/Akt and the ERK/Nrf-2 defense pathways. *Cell Death Differ*. 2010;17(7):1179–88.
  224. El Kebir D, Jozsef L, Pan W, Wang L, Petasis NA, Serhan CN, et al. 15-epi-lipoxin A4 inhibits myeloperoxidase signaling and enhances resolution of acute lung injury. *Am J Respir Crit Care Med*. 2009;180(4):311–9.
  225. Sunderland N, Hennessy A, Makris A. Animal models of pre-eclampsia. *Am J Reprod Immunol*. 2011;65(6):533–41.